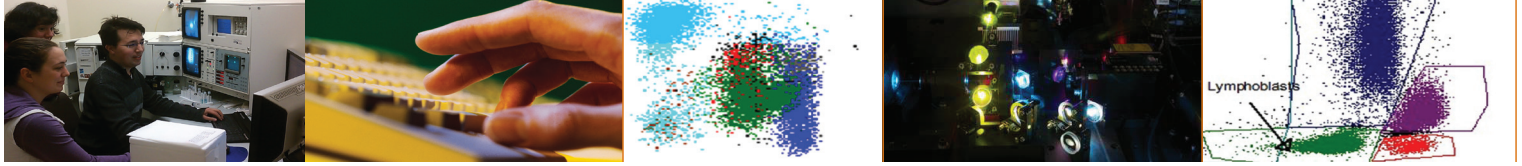


# OHSU Research Cores and Shared Resources

## Flow Cytometry

OHSU's cores are your campus technology partners dedicated to the success of your project. For optimal results, take advantage of the state-of-the-art scientific resources within the OHSU community.

[www.ohsu.edu/cores](http://www.ohsu.edu/cores)



**The OHSU Flow Cytometry Shared Resource (FCSR) has operated as a core resource for OHSU investigators since 1996 and provides advanced flow cytometry instrumentation, technical expertise and technical services.**

**Director**  
Philip Streeter Ph.D.

**Operators**  
Pamela Canaday  
Sara Christensen  
Brianna Garcia  
Dorian LaToacha

**Location**  
Marquam Hill Campus  
Richard Jones Hall  
Room 5380  
3181 SW Sam Jackson  
Park Road,  
Portland OR 97239



**Email**  
[streetep@ohsu.edu](mailto:streetep@ohsu.edu)

**Phone**  
503.418.CYTO (2986)

**Web**  
[www.ohsu.edu/flowcytometry](http://www.ohsu.edu/flowcytometry)

### Introduction

The Flow Cytometry Shared Resource (FCSR) personnel provide training in data interpretation, experiment design and routine instrument operation, offering investigators the cost-saving option of doing some of the work themselves. Resource personnel also support investigators by providing operator-assisted fluorescence-activated cell sorting, analytical flow cytometry and data analysis.

### Equipment

- DVS Sciences CyTOF
- BD LSRFortessa
- BD LSR II
- BD FACSCanto II
- BD FACSVantage cell sorter
- BD inFlux cell sorters
- MACSQuant
- Luminex 200

### Services

1. Quantitative measurement of fluorescent reporters to assess the distribution of specific molecules within cell populations.
2. Sorting to isolate purified cell populations based on detection of specific probes such as antibodies and fluorescent proteins.
3. Analysis of multiple characteristics such as relative cell size, antibody binding to cell surface or intracellular biomarkers, DNA and RNA content, and fluorescent protein expression.
4. Functional assays to measure apoptosis, enzyme activity or calcium flux.
5. Cells can be sorted into 5-15 mL conical tubes, eppendorf tubes or into multi-well plates.
6. After acquisition of data, investigators will receive data files and/or data plots. In addition, for sorting experiments, investigators will receive sorted cells and an analysis of post-sort purity.