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## Electronic supplementary material

ESM 1

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REVIEW

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# The Instrumented Fetal Sheep as a Model of Cerebral White Matter Injury in the Premature Infant

6

Stephen A. Back · Art Riddle · Justin Dean ·  
A. Roger Hohimer

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**Abstract** Despite advances in neonatal intensive care, survivors of premature birth remain highly susceptible to unique patterns of developmental brain injury that manifest as cerebral palsy and cognitive-learning disabilities. The developing brain is particularly susceptible to cerebral white matter injury related to hypoxia-ischemia. Cerebral white matter development in fetal sheep shares many anatomical and physiological similarities with humans. Thus, the fetal sheep has provided unique experimental access to the complex pathophysiological processes that contribute to injury to the human brain during successive periods in development. Recent refinements have resulted in models that replicate major features of acute and chronic human cerebral injury and have provided access to complex clinically relevant studies of cerebral blood flow and neuroimaging that are not feasible in smaller laboratory animals. Here, we focus on emerging insights and methodologies from studies in fetal sheep that have begun to define cellular and vascular factors that contribute to white matter injury. Recent advances include spatially defined measurements of cerebral blood flow in the utero, the definition of cellular maturational factors that define the topography of injury and the application of high-field magnetic resonance

imaging to define novel neuroimaging signatures for specific types of chronic white matter injury. Despite the higher costs and technical challenges of instrumented preterm fetal sheep models, they provide powerful access to clinically relevant studies that provide a more integrated analysis of the spectrum of insults that appear to contribute to cerebral injury in human preterm infants.

**Keywords** Myelin · Oligodendrocyte · White matter · Ovine · Hypoxia-ischemia · Cerebral blood flow · MRI · Cerebral palsy

## Overview of the Clinical Problems Accessible in Preterm Fetal Sheep

The last decade has seen a resurgence of interest in instrumented preterm fetal sheep preparations to study the complex pathophysiological processes that contribute to brain injury in the preterm infant. The merits of a large pre-clinical animal model have become increasingly recognized, as it has become apparent that rodent models have significant limitations to study injury to the developing human brain [1]. Not only do preterm fetal sheep preparations closely replicate major features of acute and chronic human preterm brain injury, but they also provide access to complex, clinically relevant studies of cerebral blood flow and neuroimaging that are not feasible in smaller laboratory animals.

Fetal sheep preparations have provided powerful access to large animal pre-clinical testing of neuroprotective agents for the treatment of neonatal encephalopathies related to hypoxia-ischemia or maternal fetal infection. It is increasingly recognized that the selection of appropriate animal models is critical to identify promising therapeutic agents and to reduce false negative outcomes [2]. The substantial

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65 translational advantages of studies in a large preclinical  
 66 animal model are exemplified by pre-clinical studies in near  
 67 term fetal sheep [3] that lead to the head cooling trials for  
 68 neonatal encephalopathy, and the implementation of cere-  
 69 bral hypothermia protocols in neonatal intensive care units  
 70 worldwide [4].

71 In recent years, studies of hypoxic-ischemic injury to the  
 72 developing brain have yielded an increasingly complex and  
 73 controversial set of observations related to the pathogenetic  
 74 mechanisms that result in injury to white and gray matter  
 75 structures within the neuraxis. We will focus on approaches  
 76 to study cerebral white matter injury (WMI), the major form of  
 77 brain injury, and the leading cause of chronic neurological  
 78 disability in survivors of premature birth [5]. The broader  
 79 questions of the pathogenetic mechanisms that relate WMI to  
 80 cortical and subcortical gray matter injury have been recently  
 81 reviewed [6].

82 Brain injury in preterm survivors has an unexplained pre-  
 83 dilection for cerebral white matter. The period of highest risk  
 84 for WMI is ~23 to 32 weeks postconceptional age. Although  
 85 major advances in the care of premature infants have resulted  
 86 in striking improvements in the survival of very low birth  
 87 weight infants (<1.5 kg), improved survival has been accom-  
 88 panied by a significant increase in the number of pre-term  
 89 survivors with long-term neurological deficits [7]. The major  
 90 consequences of this injury are permanent motor impairment  
 91 (i.e., cerebral palsy [CP]) ranging from mild to profound  
 92 spastic motor deficits [8–13], as well as a broad spectrum of  
 93 cognitive, social behavioral, attentional, visual, and learning  
 94 disabilities that manifest by school age in 25 to 50% of  
 95 children [14–18]. In preterm survivors, magnetic resonance  
 96 image (MRI)-defined WMI, but not gray matter injury man-  
 97 ifests in the first months of life as abnormal movements that  
 98 are predictive of CP [19–21]. The impact of WMI can be  
 99 appreciated from a recent large population-based study of  
 100 children with CP. Perinatal WMI (including periventricular  
 101 leukomalacia) was the most common finding, which was seen  
 102 in almost half (42.5%) of the affected children [22]. WMI is  
 103 not exclusively associated with prematurity and it is increas-  
 104 ingly appreciated in term infants [23–25]. Infants with com-  
 105 plex congenital heart disease are at particular risk for WMI  
 106 and delayed brain maturation [26–28]. Because very low birth  
 107 weight infants comprise approximately 1.5% of the 4 million  
 108 live births in the United States alone each year, the worldwide  
 109 social and economic burden is considerable. The average  
 110 lifetime costs per person with CP is estimated to be ~1 million  
 111 dollars in the United States. [29]. An understanding of the  
 112 cellular and molecular basis of preterm WMI is thus urgently  
 113 needed to develop effective interventions to prevent these  
 114 lifelong neurological disabilities. Hence, there continues to  
 115 be a critical need for suitable animal models that permit  
 116 hypothesis-driven studies of the pathogenesis of WMI in  
 117 human preterm survivors.

**Advantages and Disadvantages of the Fetal Sheep  
 to Model WMI** 118  
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Fetal sheep preparations require substantial cost and infra-  
 structure to support the surgical instrumentation and post-  
 operative care of large laboratory animals. Fetal sheep  
 studies require a highly skilled surgical team, a large animal  
 operating facility suitable for sterile operations, specialized  
 veterinary care, and access to reliable breeders. Despite  
 these challenges, many preparations have yielded very re-  
 producible results with low morbidity and mortality, thereby  
 limiting the number of animals required. Presently, the ovine  
 genome has not been fully sequenced and the molecular  
 tools available to study ovine brain injury are more limited  
 than in rodents. The rodent models, including transgenic  
 rodents, are ideal to provide more rapid, cost-effective ac-  
 cess to cellular and molecular mechanisms, which can be  
 validated subsequently in a large pre-clinical animal model,  
 such as the instrumented fetal sheep. 120  
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The *in utero* nature of the instrumented fetal sheep prep-  
 aration offers some practical advantages relative to other *ex*  
*utero* models with a gyrencephalic brain (e.g., the piglet and  
 nonhuman primate). To our knowledge, viable preterm fetal  
 piglet models have not been achieved. Even in neonatal pig-  
 lets, studies to date have only achieved relatively brief survival  
 [30]. Instrumentation of the preterm nonhuman primate fetus  
 carries a high risk for premature birth. Although controlled  
 preterm delivery of the nonhuman primate fetus is feasible,  
 neuropathological studies in preterm animals require ventila-  
 tor support in an intensive care setting [31]. During prolonged  
 ventilation, preterm baboons, for example, sustain disturban-  
 ces in brain growth and development that make the interpre-  
 tation of neuropathological studies more challenging [32–34].  
 A limitation of the instrumented fetal sheep preparation is that  
 animals are not studied under *ex utero* conditions where the  
 animals breathe room air. 136  
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Preterm (0.65 gestation or 95 days) fetal sheep models  
 have multiple distinct advantages relative to small fetal and  
 neonatal animals, which are limited by a lissencephalic  
 brain that does not resemble the nascent gyrencephalic  
 human cerebrum. In terms of its neurodevelopment, the  
 immature ovine brain is similar to the preterm human brain  
 between approximately 24 to 28 weeks in terms of the  
 completion of neurogenesis, the onset of cerebral sulcation,  
 and the detection of the cortical component of the auditory  
 and somatosensory evoked potentials [35–38]. The long  
 gestation of fetal sheep (145 days) allows selection of an  
 appropriate developmental stage over which brain insults  
 can be induced and evaluated. fetal sheep model 164Q3  
 165Q4

The abundance of cerebral white matter and its anatomic  
 similarities to that of the preterm infant make the fetal sheep  
 ideal for neuropathological correlation with humans [39,  
 40], whereas rodents have a paucity of cerebral white matter 166  
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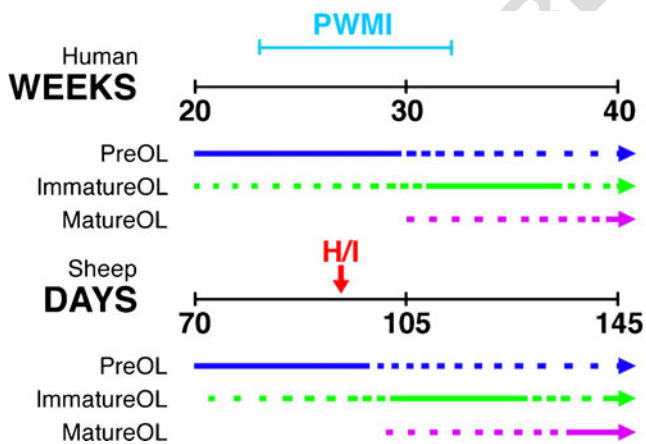


170 that differs markedly from humans. White matter maturation  
 171 in fetal sheep can be defined relative to humans through an  
 172 assessment of oligodendrocyte lineage progression and  
 173 myelination (Fig. 1). Oligodendrocyte development in the  
 174 0.65 gestation sheep fetus is similar to that of the 24- to 28-  
 175 week preterm human infant [41]. The late gestation ovine  
 176 fetus (0.9 gestation; 135 days) displays oligodendrocyte de-  
 177 velopment similar to the term human [42]. Thus, the investi-  
 178 gator can choose the appropriate developmental timing for the  
 179 insult to be evaluated. Although in both humans and sheep,  
 180 myelination begins prenatally, a notable difference is that  
 181 myelinogenesis progresses more rapidly in sheep [41] than in  
 182 humans [43], which continues to progress for months after  
 183 term birth [44]. By contrast, the rodent displays accelerated  
 184 white matter maturation and myelination that occurs entirely  
 185 postnatally [45]. In neonatal rats, for example, the progression  
 186 from human preterm equivalent to near-term equivalent oli-  
 187 godendrocyte maturation occurs rapidly during the first 5 days  
 188 after birth [44].

189 The preterm fetal sheep is susceptible to acute [41] and  
 190 chronic [46] WMI, which is very similar to humans in histo-  
 191 pathological features. The fetal sheep cerebrum has a predi-  
 192 lection for relatively selective WMI under conditions of  
 193 moderate cerebral ischemia [41], whereas rodents have a  
 194 propensity for mixed cerebral injury such that substantial gray

195 matter injury accompanies WMI [47–52]. This shortcoming,  
 196 for example, limits the relevance of rodent hypoxia-ischemia  
 197 models for the study of myelination disturbances associated  
 198 with chronic human WMI. Necrotic injury to cerebral gray  
 199 matter contributes substantially to neuro-axonal degeneration  
 200 as a cause of dysmyelination, which is not a prominent feature  
 201 of WMI in either fetal sheep [53] or contemporary human  
 202 cases of WMI [54]. The basis for this difference in suscepti-  
 203 bility to gray matter injury in rodents relative to sheep and  
 204 humans is likely to be multifactorial. The cerebrovascular  
 205 supply of rodent white matter is structurally and physiologi-  
 206 cally very dissimilar to humans and sheep [55, 56]. In addi-  
 207 tion, the timing of expression of the subtypes of glutamate  
 208 receptors appears to differ between rodents and both humans  
 209 and sheep [57–59], which may further contribute to the pro-  
 210 nounced susceptibility of neonatal rodent cerebral gray matter  
 211 to excitotoxic injury [60]. However, there are unex-  
 212 plained differences in glutamate transporter expression  
 213 between sheep and humans during preterm white matter  
 214 development where EAAT1 expression predominates in  
 215 sheep [61], whereas EAAT2 expression predominates in  
 216 humans [62].

217 The size of the preterm sheep fetus allows for chron-  
 218 ic instrumentation to enable hemodynamic measure-  
 219 ments, repeated access to blood and cerebrospinal  
 220 fluid, and chronic electrophysiological recording of the  
 221 fetal electroencephalogram [63]. In contrast to fetal  
 222 sheep preparations, the small size of rodents is a major  
 223 technical limitation for a wide range of invasive phys-  
 224 iological measurements, as well as for studies that seek  
 225 to achieve high resolution neuroimaging by using MRI  
 226 [46, 64]. Thus, it is feasible to study well-defined brain  
 227 insults in fetal sheep with reliable measurements of  
 228 blood pressure (BP), oxygenation, cerebral blood flow,  
 229 and metabolism. Chronic instrumentation also allows a  
 230 wide range of practical and clinically pertinent cerebral  
 231 insults to be administered. These include global cephalic  
 232 ischemia [65], systemic hypotension or hypoxemia  
 233 [66–68], single or repeated cord occlusion [69–71], in-  
 234 creased intracranial pressure [72], and administration of  
 235 infectious agents or exogenous inflammatory mediators  
 236 [73–75]. These insults can be graded in intensity and  
 237 duration to mimic the human situation. In each case, the  
 238 stressor can be well-described, if not regulated, by con-  
 239 ventional monitoring measurements. Thus, a wide range  
 240 of pathogenetic events can be evaluated physiologically  
 241 and correlated neuropathologically with the distribution  
 242 and extent of white matter damage. Moreover, the nat-  
 243 ural progression of various types of WMI can be evaluated in  
 244 a time frame ranging from days to weeks to months after the  
 245 insult. From a practical standpoint, the availability, cost, and  
 246 ease of breeding the sheep makes it a more practical large  
 247 animal model than the nonhuman primate. In addition, the size



**Fig. 1** Summary diagram that compares the timing of appearance of human *versus* ovine immature oligodendrocytes (O4 + O1+). The human is depicted during the latter half of gestation (20-40 weeks) and is based on data previously reported [133]. The progression of the oligodendrocyte lineage in fetal sheep white matter development has been previously described [41, 42]. The period shown (~70-145 days) roughly corresponds with the fetal human during the latter half of gestation. Solid lines indicate the developmental period when each oligodendrocyte lineage stage predominates. The dotted lines indicate the period when these stages are a minor population. Note that in many studies hypoxia-ischemia (H/I) (arrow) is administered at 90 to 95 days, which coincides with the high-risk period for periventricular white matter injury (PWMI) at ~23 to 32 weeks. preOLs are progenitors late oligodendrocytes

248 and docile nature of the sheep supports the feasibility of either  
 249 *in utero* or *ex vivo* neuroimaging studies [46, 76].

250 **Hypoxia-Ischemia in Fetal Sheep Generates Pathological**  
 251 **Features of WMI**

252 The ovine fetal sheep offers significant advantages to analyze  
 253 systemic hemodynamic disturbances that regulate cerebral  
 254 blood flow and metabolism in preterm cerebral white matter.  
 255 The sheep fetus displays cerebral hemodynamics similar to  
 256 humans and permits repeated physiological measurements *in*  
 257 *utero* in the unanesthetized state. Importantly, as the  
 258 human fetus [77–80], the fetal sheep displays a very limited  
 259 range of cerebral autoregulation under normal conditions and  
 260 a pressure-passive cerebral circulation when subjected to sys-  
 261 temic hypoxia and associated hypotension [66, 81–84]. More-  
 262 over, measurements of BP, electroencephalography, blood  
 263 oxygenation, and other vital variables can be correlated with  
 264 acute changes in cerebral blood flow and metabolism.

265 Multiple lines of evidence support a role for cerebral  
 266 ischemia in the pathogenesis of WMI in very low birth  
 267 weight human infants [5, 85, 86]. Given the limitations of  
 268 human studies to directly link blood flow disturbances with  
 269 WMI, studies in fetal sheep have greatly strengthened our  
 270 understanding of the contribution of cerebral hypoxia-  
 271 ischemia to WMI. These experimental studies support that  
 272 a complex interplay of factors related to cerebrovascular  
 273 immaturity predispose preterm cerebral white matter to in-  
 274 jury from hypoxia-ischemia. Studies of global cerebral  
 275 hypoperfusion found that the mid-gestation animal dis-  
 276 played a predilection to subcortical WMI, whereas the near  
 277 term animal displayed predominantly parasagittal cortical  
 278 neuronal injury [65, 87]. A variable degree of WMI was  
 279 also detected after systemic hypotension arising from inter-  
 280 mittent or partial umbilical cord occlusion [88, 89]. By  
 281 contrast, in the near term animal, repeated umbilical cord  
 282 occlusion produced injury to both the periventricular white  
 283 matter and the cerebral cortex [90]. Systemic hemorrhagic  
 284 hypotension in the 0.75 gestation fetus resulted in mostly  
 285 necrotic WMI with focal necrotic lesions or axonal swellings  
 286 in the periventricular white matter [91]. Preterm ovine white  
 287 matter lesions were also detected after repeated systemic fetal  
 288 endotoxin exposure that triggered both transient hypoxemia  
 289 and hypotension [92, 93]. The importance of cerebral ischemia  
 290 is supported by studies where WMI was detected only infre-  
 291 quently in models of hypoxemia in which a restriction in  
 292 uteroplacental blood flow resulted in decreased oxygen deliv-  
 293 ery and mild acidemia to the fetus without systemic hypoten-  
 294 sion or cerebral hypoperfusion [67, 94, 95]. A model of fetal  
 295 metabolic acidemia induced by maternal hypoxemia similarly  
 296 produced mild-to-moderate injury in mid-gestation and near-  
 297 term sheep [96]. Hence, cerebral hypoperfusion in conjunction

with hypoxia appears to be a critical factor to generate signifi-  
 cant WMI in the preterm fetal sheep.

Fetal sheep have also provided an important model to define  
 mechanisms of oxidative injury from cerebral hypoxia-  
 ischemia. In part, this is due to the large fetal cerebral hemi-  
 spheres, which permit blood sampling from the venous sagittal  
 sinus and placement of intracerebral probes for dialysis studies.  
 Initial studies found that reperfusion after cerebral ischemia  
 was an important source of free radical formation in the near  
 term fetal sheep brain [97]. Partial umbilical cord occlusion  
 resulted in a delayed increase in lipid peroxidation in the  
 frontal and parietal white matter in near term fetal sheep [98].  
 Dialysis studies in near term [99] and preterm [100, 101] sheep  
 have demonstrated that the enhanced generation of reactive  
 oxygen species does not occur during the immediate ischemia-  
 reperfusion period, but it is observed after a significant delay.

**Pathophysiological Mechanisms of WMI Related**  
**to the Brachiocephalic versus Carotid Occlusion Models**  
**of Global Cerebral Ischemia**

We have used 2 different approaches to achieve global cerebral  
 ischemia. The first is to place an occluder on each of the carotid  
 arteries in the neck. The second is to place a single occluder  
 around the brachiocephalic artery in the chest. The relative  
 advantages and disadvantages of each approach are discussed  
 as follows. To confine the cerebral blood supply to the carotid  
 arteries for both approaches, the occipito-vertebral anastomosis  
 (OVA) are ligated bilaterally. These anastomoses connect the  
 vertebral arteries, supplied by the thoracic aorta, with the  
 external carotid arteries that are fed by the brachiocephalic  
 [102].

The dominant factors related to the generation of WMI in  
 most, if not all, models are: 1) the content of oxygen and  
 glucose in the blood, 2) the driving or perfusion pressure  
 and hence cerebral blood flow (CBF), and 3) the duration of  
 the insult. In preterm fetal sheep, it is often feasible for these  
 important variables to be manipulated and measured, either  
 in groups of animals or in single individuals. The patho-  
 physiological disturbances associated with these factors are  
 variously weighted, depending on the model being used.

With models of maternal hypoxemia (e.g., high altitude  
 or a low inspired oxygen fraction), fetal oxygenation falls  
 while BP rises transiently, but then returns to normal, and  
 then falls below normal [67, 68, 96, 103–106]. Perinatal  
 hypoxic-ischemic encephalopathy models that involve the  
 entire body frequently use umbilical cord occlusion and are  
 accompanied by reduced systemic fetal arterial oxygen ten-  
 sion and content [70, 107, 108]. Injury in the umbilical cord  
 occlusion models can be modulated by adjusting the duration  
 of occlusion relative to the onset of systemic hypotension. In  
 cord occlusion models, BP initially increases to compensate



348 for diminished oxygenation, but as systemic and cardiac hyp- 401  
 349 oxia progressively intensify, hemodynamic and cardiac de- 402  
 350 compensation occurs with a resultant fall in BP. It appears that 403  
 351 significant brain damage occurs only when the hypoxia and 404  
 352 hypotension are allowed to progress until near death condi- 405  
 353 tions occur. However, as cardiovascular compensation fails 406  
 354 with time, substantial albeit variable reductions in BP occur. 407  
 355 The residual brain blood flow has seldom been measured, but 408  
 356 for brain injury to occur, pressures must fall below one third of 409  
 357 normal for at least 10 to 15 minutes, depending on the ani- 410  
 358 mal's age and the model being used [41, 109]. The importance 411  
 359 of the central autonomic system and adrenal stress hormones 412  
 360 remains unclear, as well as the systemic blood concentrations 413  
 361 of metabolic substrates, such as glucose and products such as 414  
 362 lactate. The fetal sheep brain, in particular when immature and 415  
 363 hypoxemic, has essentially no ability to autoregulate [81–83]. 416  
 364 Hence, in umbilical cord occlusion models, the fall in BP 417  
 365 exacerbates the fetal brain hypoxia with a consequent reduc- 418  
 366 tion in CBF that leads to partial ischemia. Thus, occlusive 419  
 367 cerebral ischemia models differ from systemic hypoxic- 420  
 368 ischemic encephalopathy models in several significant ways. 421  
 369 A major practical advantage of the occlusive models is that the 422  
 370 heart is largely unaffected. Essentially, cerebral damage can be 423  
 371 reliably generated without the significant fetal deaths and 424  
 372 morbidity that are inherently associated with other models 425  
 373 that require near death conditions to ensure brain damage. 426

374 Unlike *in utero* systemic hypoxemia or asphyxia models, 427  
 375 the models that use carotid or brachiocephalic artery (BCA) 428  
 376 occlusions cause an immediate fall in perfusion pressure and 429  
 377 CBF, and thus have a well-defined onset of the insult [41, 65, 430  
 378 110]. Cerebral ischemia in these models is generally global 431  
 379 and severe but not complete, especially when collateral circ- 432  
 380 ulation is left intact (discussed as follows). Importantly, some 433  
 381 CBF persists and the oxygenation and glucose levels of this 434  
 382 residual flow have important influence on the extent of dam- 435  
 383 age. The residual blood flow is the most difficult parameter to 436  
 384 quantify. Even in fetal sheep models, CBF is only infrequently 437  
 385 measured and BP is often used as a surrogate marker of CBF.

386 There are a number of considerations that guide the selec- 438  
 387 tion of a model that uses carotid *versus* common BCA occlu- 439  
 388 sion. However, the differences between bi-carotid and BCA 440  
 389 occlusion are not as great as the difference between either of 441  
 390 those models and cord occlusion models. The carotid arteries 442  
 391 in the 0.65 sheep fetus are small and there is a relatively greater 443  
 392 risk that commercially available occluders can inadvertently 444  
 393 obstruct flow chronically if placement and routing techniques 445  
 394 are not optimal. By contrast, the BCA is a larger and much 446  
 395 more stable vessel for the placement of a single occluder [84, 447  
 396 103]. In sheep, the BCA supplies the entire head, including the 448  
 397 carotids, as well as both axillary arteries, which supply the 449  
 398 forelegs. The BCA is the only major artery that supplies the 450  
 399 upper body. Occlusion of the BCA is similar to bilateral carotid 451  
 400 occlusion in terms of perfusion to the head and brain, but

different in that forelimb perfusion is also reduced, which does 401  
 not occur if flow to both carotids is completely restricted. 402

403 One important consequence of the BCA occlusion model 404  
 405 is that proximal BP to the rest of the fetal body and the 406  
 407 placental circulation is subject to an elevated pressure. This 408  
 409 probably results in an increase in umbilical flow that causes 410  
 411 a moderate but significant rise in arterial oxygenation. The 412  
 413 same effect probably occurs with bi-carotid occlusion, but it 414  
 415 would be expected to be smaller. Either preparation can be 416  
 417 coupled with a lowered maternal inspired oxygen fraction to 418  
 419 counteract elevations in arterial oxygenation or even generate 420  
 421 a fetal hypoxemia in addition to cerebral ischemia. 422

423 The OVA is small but potentially important in models 424  
 425 where cephalic occlusions are studied [65, 102]. They provide 426  
 427 an important anastomosis between the anterior circulation 428  
 429 provided by the carotid arteries and the posterior circulation 430  
 431 derived from the vertebral arteries. The OVAs are likely to be 432  
 433 variable in size from animal to animal, perhaps linked to 434  
 435 variations in anatomy related to their supply at or near the 436  
 437 Circle of Willis. They connect to the carotids near the lingual 438  
 439 branch, and therefore they are distal to the BCA occluder, and 440  
 441 to all but the most rostral placements of the carotid occluders. 442  
 443 Hence, they act to support brain blood flow when systemic 444  
 445 pressure is normal but BCA or carotid occlusions are used. 446

447 Hence, in both the bi-carotid and BCA occlusion models, it 448  
 449 is important to ligate the OVAs to achieve near complete 450  
 451 global cerebral ischemia. The exact amount of flow they 452  
 453 provide, especially to the preterm fetus, has not been carefully 454  
 455 determined. In normally oxygenated sheep, it is not clear 456  
 457 whether there is a net flow and if there is, then in what 458  
 459 direction. During either carotid or BCA occlusions, flow is 460  
 461 certainly from the vertebrals to the carotids distal to the site of 462  
 463 placement of the occluders. The amount of flow in the OVA, 464  
 465 while likely to be variable from animal to animal, is clearly 466  
 467 sufficient to cause variability in brain damage in the cephalic 468  
 469 ischemia models, unless they are ligated. 470

**Role of Vascular End Zones in Cerebral WMI** 437

438 The role of cerebral vascular immaturity in the pathogenesis 439  
 440 of WMI has been difficult to study in preterm infants. 441  
 442 Analysis of the vascular supply to the periventricular white 443  
 444 matter has yielded controversial results. The periventricular 445  
 446 white matter has 2 major blood supplies. Perforating arteries 447  
 448 branch from leptomeningeal arteries, penetrate the cerebral 449  
 450 cortex, and terminate as capillary beds adjacent to the ven- 451  
 452 tricles. Branches of choroidal and striate arteries project 453  
 454 toward the lateral ventricles and then deviate away from 455  
 456 the ventricle toward their final termination in vascular capil- 457  
 458 lary beds in the periventricular white matter. Although these 459  
 460 vascular beds may collectively form vascular end zones and 461  
 462 border zones that render the periventricular white matter 463

451 particularly susceptible to ischemia, physiological studies in  
 452 support of this concept are lacking. Hence, existence of these  
 453 border zones remains controversial [111–113]. The presence  
 454 of these vascular zones would provide a mechanism for WMI  
 455 based on the notion that when periventricular white matter  
 456 flow falls below a critical threshold, this region would display  
 457 greater susceptibility to WMI relative to the better-perfused  
 458 cerebral cortex.

459 To address this controversy, we developed methods to spa-  
 460 tially quantify CBF, because large differences in flow in small  
 461 regions can be obscured when averaged with larger unaffected  
 462 regions, as is the case with more global measures of cerebral  
 463 blood flow [41]. To be able to analyze CBF disturbances in  
 464 regions of periventricular white matter vulnerable to hypoxic-  
 465 ischemic injury, we developed a novel method to achieve high  
 466 resolution spatial blood flow measurements on the brains of  
 467 immature fetal sheep. This was achieved by determining the  
 468 location in the fetal brain of thousands of individual fluores-  
 469 cently labeled microspheres [41, 109]. This high resolution  
 470 blood flow technique is illustrated in Fig. 2. Basal or control  
 471 blood flow in 2 of 16 “virtual” 2-mm thick coronal sections of  
 472 a 0.65 gestation fetal sheep brain are shown. The image in the  
 473 upper left represents a three-dimensional reconstruction of the  
 474 approximately 1000 Imaging CryoMicrotome images (Barlow  
 475 Scientific, Olympia, WA) obtained to define the location of  
 476 each microsphere. Two coronal two-dimensional virtual sec-  
 477 tions, at the level of the (image 1) parietal and (image 2) frontal  
 478 white matter (Fig. 2), where white matter damage was ob-  
 479 served, show the two-dimensional representation of blood flow  
 480 under basal conditions. Blood flow is displayed as a “con-  
 481 volved” image of the density of microsphere distribution and  
 482 quantified (ml/min) in the pseudocolor scale shown. Basal  
 483 blood flow was particularly high, for example, in the thalamic

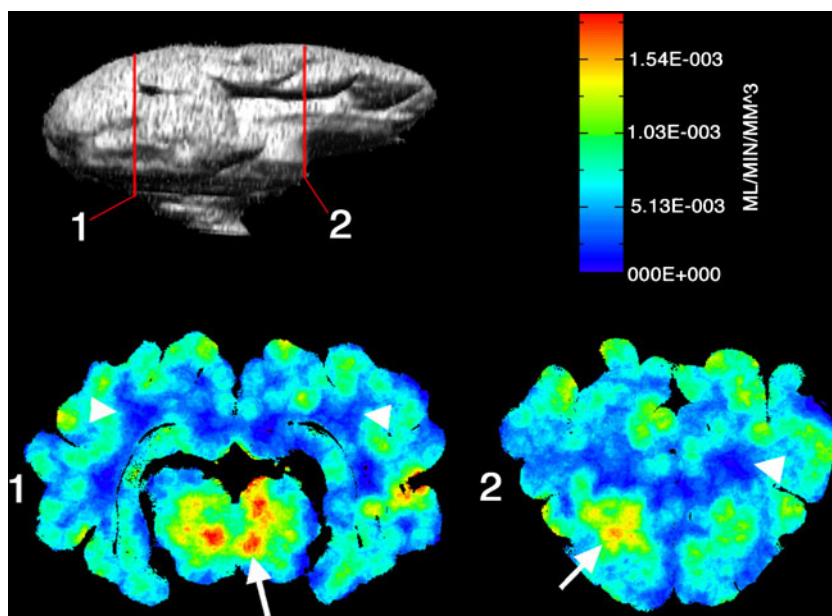
nuclear groups in the midbrain (Fig. 2, image 1, arrow) relative  
 to the parietal cortex. By contrast, the basal blood flow of the  
 parietal and frontal periventricular white matter was markedly  
 lower (Fig. 2, arrowheads). Cortical blood flow is much higher  
 than periventricular white matter flow. This degree of spatial  
 resolution can only be achieved in brains at least the size of the  
 immature fetal sheep.

To seek evidence for vascular end and border zones in fetal  
 cerebral white matter, we measured blood flow in histopatho-  
 logically defined regions of injury in cerebral cortex and white  
 matter in preterm fetal sheep [109]. Although white matter  
 blood flow was lower than cerebral gray matter, there was no  
 evidence for pathologically significant gradients of fetal blood  
 flow within the periventricular white matter under conditions of  
 global partial ischemia or reperfusion. White matter lesions did  
 not localize to regions susceptible to greater ischemia, nor did  
 less vulnerable regions of cerebral white matter have greater  
 flow during ischemia. An alternative explanation for the topog-  
 raphy of cerebral white matter lesions is the distribution of  
 susceptible cell types (described as follows), particularly late  
 oligodendrocyte (OLs) progenitors (preOLs) that are particu-  
 larly susceptible to hypoxia-ischemia.

**Relative Contributions of Hypoxia-Ischemia and OL  
 Lineage Immaturity to Acute WMI**

The preterm fetal sheep (0.65 gestation) displays heterogeneous  
 OL lineage maturation in frontal periventricular white matter  
 [41], which allowed us to define the relative contributions of  
 oligodendroglial maturational factors and vascular factors to  
 acute WMI. OL lineage maturation in medial periventricular  
 white matter (PVWM) was similar to that of a human (~23–

**Fig. 2** Quantification of fetal cerebral blood flow *in situ* under conditions of basal flow. The top image represents a three-dimensional surface reconstruction (Volocity, Improvion, Lexington, MA) of fluorescence images of a 0.65 gestation ovine control brain that indicates the frontal and parietal levels to which the lower blood flow images correspond. Representative grayscale basal flow images show higher blood flow (arrows) in the pons (image 1) and subcortical gray matter (image 2), and lower flow (dark gray) in the periventricular white matter (PVWM) (arrowheads)



514 28 weeks gestation) in that preOLs were the major OL stage  
 515 present. By contrast, lateral PVWM was more differentiated  
 516 and contained predominantly pre-myelinating and early myeli-  
 517 nating immature OLs. Surprisingly, we found that moderate  
 518 cerebral ischemia did not uniformly damage the PVWM. The  
 519 medial and lateral PVWM sustained differing degrees of acute  
 520 injury, even though they sustained a similar degree of low flow  
 521 during prolonged ischemia-reperfusion. Hence, while global  
 522 ischemia was necessary for WMI, no regional differences in  
 523 blood flow were found within the PVWM under basal or  
 524 ischemic conditions to account for the differences in cell death  
 525 between medial and lateral PVWM. Rather, differences in the  
 526 topography of WMI were closely correlated with the distribu-  
 527 tion of vulnerable preOLs. Interestingly, in regions of preOL  
 528 degeneration, other neural cell types (astrocytes, microglia, and  
 529 axons) were markedly more resistant to injury.

530 In a fetal rabbit model of placental insufficiency, signif-  
 531 icant global fetal hypoxia-ischemia caused minimal WMI at  
 532 fetal day 22, but a similar insult 3 days later in gestation  
 533 caused pronounced WMI [114]. The relative susceptibility  
 534 of the white matter at these 2 developmental ages coincided  
 535 with the timing of appearance of susceptible preOLs. Taken  
 536 together, these findings suggest that perturbations in cere-  
 537 bral blood flow are necessary, but not sufficient to explain  
 538 the distribution of WMI. The developmental predilection for  
 539 WMI appears to be related to both the timing of appearance  
 540 and regional distribution of susceptible preOLs.

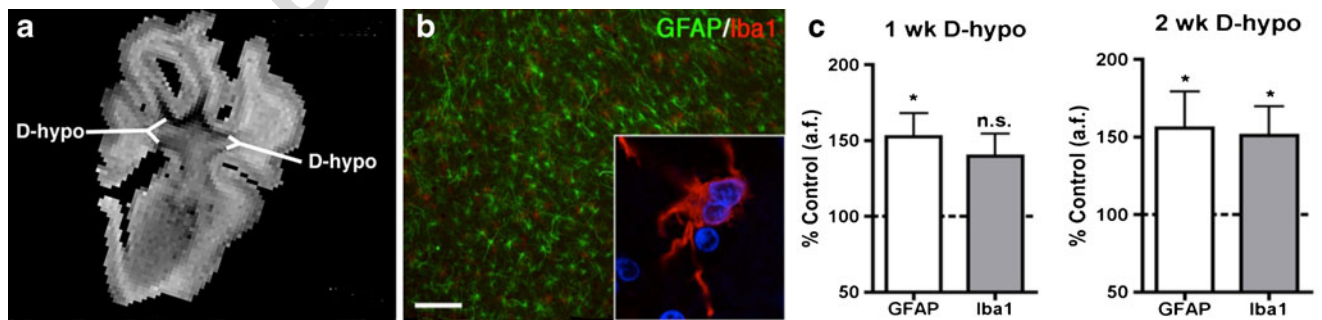
541 **Role of High-Field MRI to Define Pathological Features**  
 542 **of WMI**

543 Although MRI is the optimal imaging modality to define  
 544 WMI in preterm survivors [115–118], the histopathological  
 545 features of MRI signal abnormalities have been mostly

defined for WMI where periventricular leukomalacia predom- 546  
 inates [64, 119–123]. With the pronounced shift to milder 547  
 forms of human WMI defined by quantitative and diffusion- 548  
 weighted MRI, there is a need to define the cellular features of 549  
 these lesions. We have recently analyzed diffuse lesions in a 550  
 preterm fetal sheep model where animals survived for 1 or 551  
 2 weeks after global cerebral ischemia. This preparation gen- 552  
 erated a spectrum of WMI very similar to that observed from 553  
 human autopsy studies, as well as a reduction in cerebral WM 554  
 volume similar to that observed in preterm survivors 555  
 [116–118]. We developed registration algorithms to analyze 556  
 the histopathological features of 3 classes of MRI-defined 557  
 lesions identified by *ex vivo* imaging at high field (12 Tesla) 558  
 [46]. Each lesion type displayed unique astroglial and micro- 559  
 glial responses that corresponded to distinct forms of necrotic 560  
 or non-necrotic WMI. 561

The major type of lesion identified by MRI was a novel 562  
 diffuse hypointense signal abnormality (D-hypo) identified on 563  
 T2-weighted images at 1 and 2 weeks after global cerebral 564  
 ischemia (Fig. 3A). The physical-chemical basis for these 565  
 highly hypointense lesions is unclear, but they did not corre- 566  
 spond to hemorrhage. These lesions comprised 89% of total 567  
 lesion volume at 1 week and corresponded to histopathologically 568  
 defined lesions that were highly enriched in reactive astrocytes, 569  
 but were only modestly enriched in activated microglia at 570  
 weeks (Fig. 3B, C). This diffuse hypointense signal abnormal- 571  
 ity, thus appears to be a unique MRI signature for diffuse gliotic 572  
 lesions dominated by reactive astrogliosis. Importantly, large 573  
 lesions, which comprised 97% of the WMI identified, were 574  
 detected with a sensitivity of 100% at 1 week and 75% at 575  
 2 weeks. 576

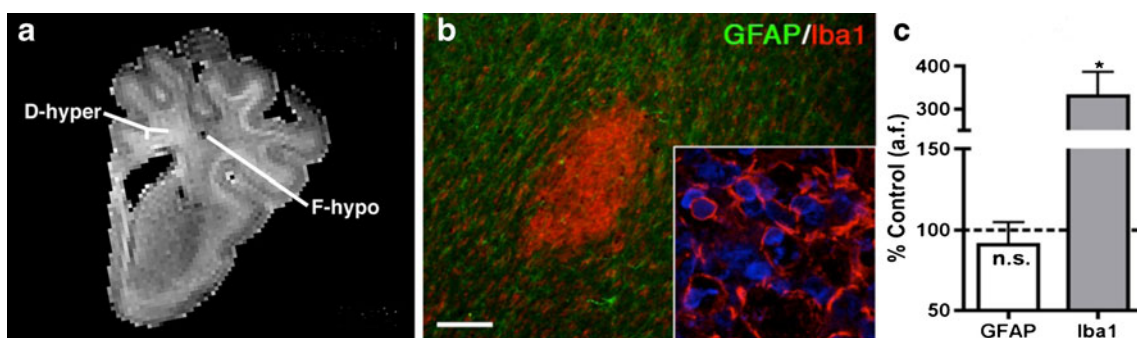
With high-field MRI, we also identified a second type of 577  
 novel focal hypointense signal abnormality (F-hypo) on T2- 578  
 weighted images at 2 weeks after global cerebral ischemia 579  
 (Fig. 4A). Although these lesions were observed in 50% of 580



**Fig. 3** Progressive histopathological and high-field magnetic resonance image (MRI)-defined features of diffuse cerebral white matter gliosis in chronic lesions generated in the 0.65 gestation ovine brain at 1 or 2 weeks after global cerebral ischemia. (A) Representative appearance and distribution of diffuse hypointense (D-hypo) lesions seen on a T<sub>2</sub>w image at 1 week after injury. (B) Diffuse white matter injury (WMI) had pronounced astrogliosis defined by immunohistochemical staining of reactive astrocytes with glial fibrillary acidic protein

(GFAP) (green) and a lesser population of Iba1-labeled microglia/macrophages (red) with a reactive morphology (inset). Nuclei in the inset are visualized with Hoechst 33342 (blue). Bar=100 μm. (C) Quantification of GFAP-labeled astrocytes and Iba1-labeled microglia within MRI-defined white matter signal abnormalities at 1 and 2 weeks after global ischemia. The D-hypo lesions had significantly elevated GFAP, consistent with a diffuse astrogliotic response to injury. \**p* < 0.05; n.s. = not significant





**Fig. 4** Histopathological and high-field magnetic resonance image (MRI)-defined features of microscopic necrosis detected in fetal ovine cerebral white matter at 2 weeks after global cerebral ischemia. (A) Representative appearance of a focal hypointense (F-hypo) lesion seen on a T<sub>2</sub>w image at 2 weeks after injury. Note the substantial difference in the F-hypo lesion relative to a diffuse gliotic lesion at 2 weeks, which appears more hyperintense (D-hyper). (B) A typical microscopic

necrotic lesion defined by a discrete focus of immunohistochemical staining for reactive microglia and macrophages with Iba1 (red; inset) and a paucity of staining for astrocytes with glial fibrillary acidic protein (GFAP) (green). Nuclei in the inset are visualized with Hoechst 33342 (blue). Bar=100 μm. (C) F-hypo lesions had markedly increased Iba1 labeling and no significant difference in GFAP labeling versus control. \**p*<0.05; n.s. = not significant

581 the animals, they only comprised ~1.5% of total lesion volume  
 582 and corresponded to histopathologically defined lesions that were  
 583 highly enriched in reactive microglia and macrophages, but  
 584 contained a paucity of astrocytes (Fig. 3B, C). Thus, this focal  
 585 hypointense signal abnormality corresponded to small discrete  
 586 regions of focal microscopic necrosis that were detected at sub-  
 587 millimeter resolution. Both the incidence and lesion burden of  
 588 these small necrotic foci of WMI were quite similar to that  
 589 observed in human WMI [54]. Although these lesions are  
 590 readily detected by histopathology, they are not readily detected  
 591 by MRI at lower field strength. Hence, high-field MRI provides  
 592 a unique signature for microscopic necrosis.

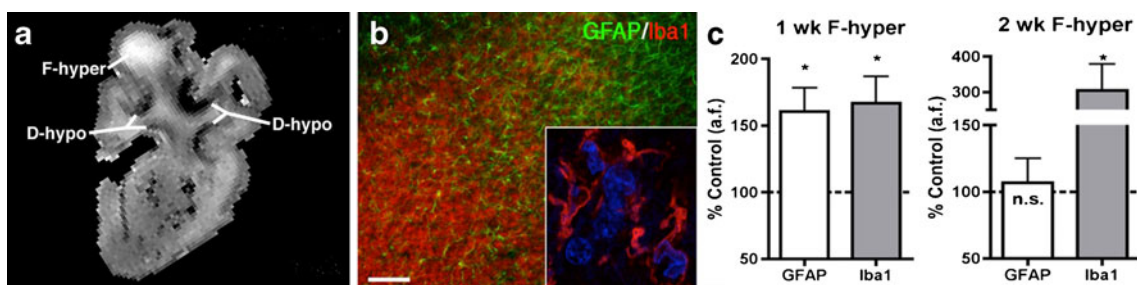
in reactive microglia and macrophages (Fig. 5B, C) and features  
 of acute axonal degeneration, including dystrophic axons and  
 axonal spheroids (not shown). Thus, these lesions displayed the  
 characteristic features of a diffuse macroscopic necrotic lesion.

These *ex vivo* high-field MRI studies suggest that current  
 clinical MRI field strength may be a limiting factor to detect  
 diffuse gliosis, as well as microscopic necrosis (discussed as  
 follows). Additional clinical pathological studies in fetal sheep  
 or other relevant models are needed to determine whether high-  
 field MRI can provide greater sensitivity to identify diffuse  
 WMI than what is currently feasible at lower field strengths.

593 A third class of lesions detected on T2-weighted images by  
 594 high-field MRI were focal hyperintense lesions (F-hyper)  
 595 (Fig. 5A). These lesions also had low values for fractional  
 596 anisotropy and high apparent diffusion coefficient values, con-  
 597 sistent with high water content. In fact, the F-hyper  
 598 lesions were defined histologically by a progressive loss of  
 599 astrocytes between 1 and 2 weeks and a marked enrichment

### MRI-Guided Ultrastructural Studies of Axonal Degeneration in WMI

Axonal degeneration is a common feature of necrotic WMI  
 and can be detected both within and distal to large necrotic  
 foci [124]. Whether primary axonal injury occurs in cases of



**Fig. 5** Histopathological and high-field magnetic resonance image (MRI)-defined features of diffuse macroscopic necrosis detected in fetal ovine cerebral white matter at 1 and 2 weeks after global cerebral ischemia. (A) Representative appearance from the largest focal hyperintense (F-hyper) lesion seen on a T<sub>2</sub>w image at 1 week after injury. These lesions typically localized to subcortical white matter. Note the substantial difference in the F-hyper lesion relative to the diffuse gliotic lesions, which appears much more hypointense (D-hypo). (B) A

typical macroscopic necrotic lesion defined by diffuse dense staining for reactive microglia and macrophages with Iba1 (red; inset) and a paucity of GFAP-labeled astrocytes. Nuclei in the inset are visualized with Hoechst 33342 (blue). Bar=100 μm. (C) F-hyper lesions displayed a progressive decrease in glial fibrillary acidic protein (GFAP) staining and markedly increased Iba1 labeling for microglia by 2 weeks after global ischemia. \**p*<0.05; n.s. = not significant

616 diffuse WMI that lack significant necrosis has received  
 617 limited study. During the acute phase of WMI, after global  
 618 hypoxia-ischemia, fetal sheep that lacked necrotic WMI did  
 619 not show evidence of axonal degeneration [41]. We used  
 620 quantitative electron microscopy studies in our preterm fetal  
 621 sheep model of global cerebral ischemia to define the extent  
 622 of axonal degeneration in MRI-defined diffuse WMI [53].  
 623 During the chronic phase of WMI from this same fetal sheep  
 624 preparation [46] and human [54], microscopic foci of ne-  
 625 crosis were observed that were rich in microglia, but lacked  
 626 astrocytes or axons. However, no significant axonal degen-  
 627 eration, axonal loss or shift in the distribution of axon  
 628 calibers was observed by quantitative electron microscopy  
 629 studies in preterm fetal sheep [53]. Hence, the contribution  
 630 of microscopic necrosis to axonal loss with secondary mye-  
 631 lination failure appears to be low, but there is a need for  
 632 further human neuropathological studies with sensitive  
 633 markers of axonal injury that are applied to cases of diffuse  
 634 WMI that lack necrosis.

635 **Final Conclusions**

636 The preterm human infant displays unique patterns of cerebral  
 637 injury that can now be closely replicated in the preterm fetal  
 638 sheep. Despite the higher costs and technical challenges of  
 639 preterm fetal sheep models, they provide powerful access to  
 640 pre-clinical questions related to the pathophysiology of WMI.  
 641 Recent advances include spatially defined measurements of  
 642 cerebral blood flow *in utero*, the definition of cellular matu-  
 643 rational factors that define the topography of WMI, and the  
 644 application of high-field neuroimaging to define MRI signa-  
 645 tures for specific types of chronic WMI. There is a critical  
 646 need to further define the cellular and molecular mechanisms  
 647 that mediate the progression of cerebral white and gray matter  
 648 injury. Such information is essential for the rationale design of  
 649 therapies targeted to block the initial or secondary phases of  
 650 injury, and to promote regeneration and repair during the  
 651 chronic phase.

652 With few exceptions, most studies have focused on models  
 653 of isolated exposure to cerebral hypoxia-ischemia or maternal  
 654 fetal infection. Future models are needed that more closely  
 655 reproduce the spectrum of insults that appear to contribute to  
 656 cerebral injury in human preterm infants. For example, there  
 657 are potential limitations of studies of healthy animals that do  
 658 not fully address the impact of clinically common complica-  
 659 tions, which influence the response to a single episode of  
 660 hypoxia-ischemia. These include the influence of antecedent  
 661 recurrent transient or prolonged hypoxia-ischemia, placental  
 662 insufficiency, or chronic *in utero* ischemia. Depending on the  
 663 timing of the insult, fetal infection has the potential to either  
 664 exacerbate ischemic injury or protect against it via mecha-  
 665 nisms that may involve ischemic tolerance. Elegant studies in

neonatal rodents have not been sufficiently replicated in large  
 preterm models [125–127]. To date there have been virtually  
 no neurobehavioral studies of preterm fetal sheep that  
 have sustained WMI. Greater clinical relevance of fetal sheep  
 models of WMI also may be achieved by addressing common  
 confounders of neonatal care that include the influence of  
 nutritional status, stressors, painful exposures, and recurrent  
 sedative and anesthetic exposure [128–132]. All are likely to  
 adversely influence neonatal brain development and the im-  
 pact of WMI on subsequent long-term motor and cognitive  
 development via mechanisms that are currently not well  
 understood.

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**References**

1. Ferriero DM. Can we define the pathogenesis of human periven-  
 tricular white-matter injury using animal models? *J Child Neurol*  
 2006;21:580-581.
2. Gressens P, Le Verche V, Fraser M, et al. Pitfalls in the quest of  
 neuroprotectants for the perinatal brain. *Dev Neurosci* 2011;33:189-  
 198.
3. Gunn AJ, Battin M, Gluckman PD, Gunn TR, Bennet L. Therapeutic  
 hypothermia: from lab to NICU. *J Perinat Med* 2005;33:340-346.
4. Higgins RD, Raju T, Edwards AD, et al. Hypothermia and other  
 treatment options for neonatal encephalopathy: an executive sum-  
 mary of the Eunice Kennedy Shriver NICHD workshop. *J Pediatr*  
 2011;159:851-858.
5. Volpe JJ. *Neurology of the Newborn*. Philadelphia: W.B. Saunders,  
 2008.
6. Back S. Mechanisms of Acute and Chronic Brain Injury in the  
 Preterm Infant. In: Miller S, Shevell M, eds. *Acquired Brain  
 Injury*. London: Mac Keith Press, 2012.
7. Wilson-Costello D, Friededman H, Minich N, Fanaroff A, Hack  
 M. Improved survival rates with increased neurodevelopmental  
 disability for extremely low birth weight infants in the 1990s.  
*Pediatrics* 2005;115:997-1003.
8. Hack M, Taylor H, Drotar D, et al. Chronic conditions, functional  
 limitations, and special health care needs of school-aged children  
 born with extremely low-birth-weight in the 1990's. *JAMA*  
 2005;294:318-325.
9. Miller SP, Ferriero DM, Leonard C, et al. Early brain injury in  
 premature newborns detected with magnetic resonance imaging is  
 associated with adverse neurodevelopmental outcome. *J Pediatr*  
 2005;147:609-616.
10. Beaino G, Khoshnood B, Kaminski M, et al. Predictors of cere-  
 bral palsy in very preterm infants: the EPIPAGE prospective  
 population-based cohort study. *Dev Med Child Neurol* 2010;52:  
 e119-e125.
11. Mercier CE, Dunn MS, Ferrelli KR, Howard DB, Soll RF. Neuro-  
 developmental outcome of extremely low birth weight infants

724 from the Vermont Oxford network: 1998-2003. Neonatology 725 2010;97:329-338.

726 12. Liu J, Li J, Qin GL, Chen YH, Wang Q. Periventricular leuko- 727 malacia in premature infants in mainland China. Am J Perinatol 728 2008;25:535-540.

729 13. Spittle AJ, Cheong J, Doyle LW, et al. Neonatal white matter 730 abnormality predicts childhood motor impairment in very preterm 731 children. Dev Med Child Neurol 2011;53:1000-1006.

732 14. Litt J, Taylor H, Klein N, Hack M. Learning disabilities in children 733 with very low birthweight: prevalence, neuropsychological correlates 734 and educational interventions. J Learn Disabil 2005;8:130-141.

735 15. Jacobson LK, Dutton GN. Periventricular leukomalacia: an im- 736 portant cause of visual and ocular motility dysfunction in chil- 737 dren. Surv Ophthalmol 2000;45:1-13.

738 16. Glass HC, Fujimoto S, Ceppi-Cozzio C, et al. White-matter injury 739 is associated with impaired gaze in premature infants. Pediatr 740 Neurol 2008;38:10-15.

741 17. Soria-Pastor S, Gimenez M, Narberhaus A, et al. Patterns of 742 cerebral white matter damage and cognitive impairment in ad- 743 lescents born very preterm. Int J Dev Neurosci 2008;26:647-654.

744 18. Anderson PJ, De Luca CR, Hutchinson E, Spencer-Smith MM, 745 Roberts G, Doyle LW. Attention problems in a representative 746 sample of extremely preterm/extremely low birth weight children. 747 Dev Neuropsychol 2011;36:57-73.

748 19. Constantinou JC, Adamson-Macedo EN, Mirmiran M, Fleisher BE. 749 Movement, imaging and neurobehavioral assessment as predictors 750 of cerebral palsy in preterm infants. J Perinatol 2007;27:225-229.

751 20. Spittle AJ, Brown NC, Doyle LW, et al. Quality of general move- 752 ments is related to white matter pathology in very preterm infants. 753 Pediatrics 2008;121(5):e1184-e1189.

754 21. Spittle AJ, Boyd RN, Inder TE, Doyle LW. Predicting motor 755 development in very preterm infants at 12 months' corrected 756 age: the role of qualitative magnetic resonance imaging and 757 general movements assessments. Pediatrics 2009;123:512-517.

758 22. Bax M, Tydeman C, Flodmark O. Clinical and MRI correlates of 759 cerebral palsy: the European Cerebral Palsy Study. JAMA 760 2006;296:1602-1608.

761 23. Pagliano E, Fedrizzi E, Erbetta A, et al. Cognitive profiles and 762 visuo-perceptual abilities in preterm and term spastic diplegic children 763 with periventricular leukomalacia. J Child Neurol 2007;22:282-288.

764 24. Li AM, Chau V, Poskitt KJ, et al. White matter injury in term 765 newborns with neonatal encephalopathy. Pediatr Res 2009;65:85-89.

766 25. Lasry O, Shevell MI, Dagenais L. Cross-sectional comparison 767 of periventricular leukomalacia in preterm and term children. 768 Neurology 2010;74:1386-1391.

769 26. Wernovsky G, Shillingford A, Gaynor J. Central nervous system 770 outcomes in children with complex congenital heart disease. Curr 771 Opin Cardiol 2005;20:94-99.

772 27. Miller S, McQuillen P, Hamrick S, et al. Abnormal brain devel- 773 opment in newborns with congenital heart disease. N Engl J Med 774 2007;357:1971-1973.

775 28. Licht DJ, Shera DM, Clancy RR, et al. Brain maturation is 776 delayed in infants with complex congenital heart defects. J 777 Thorac Cardiovasc Surg 2009;137:529-537.

778 29. Centers for Disease Control and Prevention. Economic costs 779 associated with mental retardation, cerebral palsy, hearing loss 780 and vision impairment — United States, 2003. MMWR Morb 781 Mortal Wkly Rep 2004;53:57-59.

782 30. Thoresen M, Haaland K, Loberg EM, et al. A piglet survival model 783 of posthypoxic encephalopathy. Pediatr Res 1996;40:738-748.

784 31. Griffith JL, Shimony JS, Cousins SA, et al. MR imaging corre- 785 lates of white-matter pathology in a preterm baboon model. 786 Pediatr Res 2012;71:185-191.

787 32. Loeliger M, Inder T, Cain S, et al. Cerebral outcomes in a preterm 788 baboon model of early *versus* delayed nasal continuous positive 789 airway pressure. Pediatrics 2006;118:1640-1653.

33. Loeliger M, Inder TE, Shields A, et al. High-frequency oscilla- 790 tory ventilation is not associated with increased risk of neuropa- 791 thology compared with positive pressure ventilation: a preterm 792 primate model. Pediatr Res 2009;66:545-550. 793

34. Verney C, Rees S, Biran V, Thompson M, Inder T, Gressens P. 794 Neuronal damage in the preterm baboon: impact of the mode of 795 ventilatory support. J Neuropathol Exp Neurol 2010;69:473-482. 796

35. Barlow R. The foetal sheep: morphogenesis of the nervous sys- 797 tem and histochemical aspects of myelination. J Comp Neurol 798 1969;135:249-262. 799

36. Bernhared C, Kolmodin G, Meyerson B. On the prenatal devel- 800 opment of function and structure in the somesthetic cortex of the 801 sheep. Prog Brain Res 1967;2:60-77. 802

37. Cook C, Gluckman P, Johnston B, Williams C. The development 803 of the somatosensory evoked potential in the unanaesthetized 804 fetal lamb. J Dev Physiol 1987;9:441-456. 805

38. Cook C, Williams C, Gluckman P. Brainstem auditory evoked 806 potential in the fetal lamb, in utero. J Dev Physiol 1987;9:429-440. 807

39. Gluckman P, Parsons Y. Stereotaxic method and atlas for the 808 ovine fetal forebrain. J Dev Physiol 1983;5:101-128. 809

40. Vanderwolf C, Cooley R. The sheep brain: a photographic series, 810 2nd ed. London, Ontario, Canada: A. J. Kirby Co., 1990. 811

41. Riddle A, Luo N, Manese M, et al. Spatial heterogeneity in 812 oligodendrocyte lineage maturation and not cerebral blood flow 813 predicts fetal ovine periventricular white matter injury. J Neurosci 814 2006;26:3045-3055. 815

42. Back SA, Riddle A, Hohimer AR. Role of instrumented fetal 816 sheep preparations in defining the pathogenesis of human peri- 817 ventricular white matter injury. J Child Neurol 2006;21:582- 818 589. 819

43. Back SA, Luo NL, Borenstein NS, Volpe JJ, Kinney HC. 820 Arrested oligodendrocyte lineage progression during human ce- 821 rebral white matter development: dissociation between the timing 822 of progenitor differentiation and myelinogenesis. J Neuropathol 823 Exp Neurol 2002;61:197-211. 824

44. Dean J, Moravec M, Grafe M, et al. Strain-specific differences in 825 perinatal rodent oligodendrocyte lineage progression and its cor- 826 relation with human. Dev Neurosci 2011;xx:xx-xx. 827

45. Craig A, Luo NL, Beardsley DJ, et al. Quantitative analysis of 828 perinatal rodent oligodendrocyte lineage progression and its cor- 829 relation with human. Exp Neurol 2003;181:231-240. 830

46. Riddle A, Dean J, JR B, et al. Histopathological correlates of 831 MRI-defined chronic perinatal white matter injury. Ann Neurol 832 2011;xx:xx-xx. 833

47. Segovia K, Mccormick M, Moravec M, et al. Arrested oligodendro- 834 cyte lineage maturation in chronic perinatal white matter injury. 835 Ann Neurol 2008;63:517-526. 836

48. Back SA, Han BH, Luo NL, et al. Selective vulnerability of late 837 oligodendrocyte progenitors to hypoxia-ischemia. J Neurosci 838 2002;22:455-463. 839

49. Follet PL, Rosenberg PA, Volpe JJ, Jensen FE. NBQX attenuates 840 excitotoxic injury to the developing white matter. J Neurosci 841 2000;20:9235-9241. 842


50. Uehara H, Yoshioka H, Kawase S, et al. A new model of white 843 matter injury in neonatal rats with bilateral carotid artery occlusion. 844 Brain Res 1999;837:213-220. 845

51. Olivier P, Baud O, Evrard P, Gressens P, Verney C. Prenatal 846 ischemia and white matter damage in rats. J Neuropathol Exp 847 Neurol 2005;64:9d98-1006. 848

52. Marret S, Mukendi R, Gadisseux J-F, Gressens P, Evrard P. Effect 849 of ibotenate on brain development: an excitotoxic mouse model 850 of microgyria and posthypoxic-like lesions. J Neuropathol Exp 851 Neurol 1995;xx:xx-xx. 852

53. Riddle A, Maire J, Gong X, et al. Differential susceptibility to 853 axonopathy in necrotic and non-necrotic perinatal white matter 854 injury. Stroke 2012;43:178-184. 855



856  Buser J, Maire J, Riddle A, et al. Arrested preOLigodendrocyte  
857 maturation contributes to myelination failure in premature  
858 infants. *Ann Neurol* 2012;71:93-109.

859 55. Vannucci RC, Lyons DT, Vasta F. Regional cerebral blood flow  
860 during hypoxia-ischemia in immature rats. *Stroke* 1988;19:245-  
861 250.

862 56. Vannucci RC, Vannucci SJ. Perinatal hypoxic-ischemic brain damage:  
863 evolution of an animal model. *Dev Neurosci* 2005;27:81-86.

864 57. Talos D, Fishman R, Park H, et al. Developmental regulation of  
865 alpha-amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid receptor  
866 subunit expression in forebrain and relationship to regional  
867 susceptibility to hypoxic/ischemic injury. I. Rodent cerebral white  
868 matter and cortex. *J Comp Neurol* 2006;497:42-60.

869 58. Talos DM, Follett PL, Folkerth RD, et al. Developmental regulation  
870 of alpha-amino-3-hydroxy-5-methyl-4-isoxazole-propionic  
871 acid receptor subunit expression in forebrain and relationship to  
872 regional susceptibility to hypoxic/ischemic injury. II. Human  
873 cerebral white matter and cortex. *J Comp Neurol* 2006;497:61-  
874 77.

875 59. Dean JM, Fraser M, Shelling AN, et al. Ontogeny of AMPA and  
876 NMDA receptor gene expression in the developing sheep white  
877 matter and cerebral cortex. *Brain Res Mol Brain Res*  
878 2005;139:242-250.

879 60. Johnston MV. Excitotoxicity in neonatal hypoxia. *MRDD Res*  
880 2001;7:229-234.

881 61. Northington FJ, Traystman RJ, Koehler RC, Martin LJ. GLT1,  
882 glial glutamate transporter, is transiently expressed in neurons  
883 and develops astrocyte specificity only after midgestation in the  
884 ovine fetal brain. *J Neurobiol* 1999;39:515-526.

885 62. Desilva TM, Kinney HC, Borenstein NS, et al. The glutamate  
886 transporter EAAT2 is transiently expressed in developing human  
887 cerebral white matter. *J Comp Neurol* 2007;501:879-890.

888 63. Gunn AJ, Bennet L. Fetal hypoxia insults and patterns of brain  
889 injury: insights from animal models. *Clin Perinatol* 2009;36:579-  
890 593.

891 64. Lodygensky G, West T, Moravec M, et al. Diffusion character-  
892 istics associated with neuronal injury and glial activation following  
893 hypoxia-ischemia in the immature brain. *Magn Reson Med*  
894 2011;66:839-835.



895 65. Reddy K, Mallard C, Guan J, et al. Maturation change in the  
896 cortical response to hypoperfusion injury in the fetal sheep.  
897 *Pediatr Res* 1998;43:674-682.

898 66. Szymonowicz W, Walker A, Yu V, Stewart M, Cannata J, Cussen  
899 L. Regional cerebral blood flow after hemorrhagic hypotension in  
900 the preterm, near-term, and newborn lamb. *Pediatr Res*  
901 1990;28:361-366.

902 67. Rees S, Breen S, Loeliger M, McCrabb G, Harding R. Hypox-  
903 emia near mid-gestation has long-term effects on fetal brain  
904 development. *J Neuropathol Exp Neurol* 1999;58:932-945.

905 68. Gleason CA, Hamm C, Jones MD Jr. Effect of acute hypoxemia  
906 on brain blood flow and oxygen metabolism in immature fetal  
907 sheep. *Am J Physiol* 1990;258(4 pt 2):H1064-H1069.

908 69. Fadlkowski A, Hammond R, Han V, Richardson B. Apoptosis in the  
909 preterm and near term ovine fetal brain and the effect of intermittent  
910 umbilical cord occlusion. *Dev Brain Res* 2002;136:165-173.

911  70. Bennet L, Rossenrode S, Gunning MI, Gluckman PD, Gunn AJ.  
912 The cardiovascular and cerebrovascular responses of the imma-  
913 ture fetal sheep to acute umbilical cord occlusion.  1999;517  
914 (pt 1):247-257.

915 71. Welin AK, Svedin P, Lapatto R, et al. Melatonin reduces in-  
916 flammation and cell death in white matter in the mid-gestation  
917 fetal sheep following umbilical cord occlusion. *Pediatr Res*  
918 2007;61:153-158.

919 72. Harris AP, Koehler RC, Gleason CA, Jones MD Jr., Traystman  
920 RJ. Cerebral and peripheral circulatory responses to intracranial  
921 hypertension in fetal sheep. *Circ Res* 1989;64:991-1000.

73. Rees S, Hale N, De Matteo R, et al. Erythropoietin is neuro- 922  
protective in a preterm ovine model of endotoxin-induced brain 923  
injury. *J Neuropathol Exp Neurol* 2010;69:306-319. 924

74. Duncan J, Cock M, Scheerlinck J, et al. White matter injury after 925  
repeated endotoxin exposure in the preterm ovine fetus. *Ped Res* 926  
2006;52:941-949. 927

75. Dean J, van de Looij Y, Sizonenko S, et al. Delayed cortical 928  
impairment following lipopolysaccharide exposure in preterm 929  
fetal sheep. *Ann Neurol* 2011;70:846-856. 930

76. Sorensen A, Pedersen M, Tietze A, Ottosen L, Duus L, Uldbjerg 931  
N. BOLD MRI in sheep fetuses: a non-invasive method for 932  
measuring changes in tissue oxygenation. *Ultrasound Obstet* 933  
*Gynecol* 2009;34:687-692. 934

77. Pyrds O. Control of cerebral circulation in the high-risk neonate. 935  
*Ann Neurol* 1991;30:321-329. 936

78. Menke J, Michel E, Hildebrand S. Cross-spectral analysis of 937  
cerebral autoregulation dynamics in high risk preterm infants 938  
during the perinatal period. *Pediatr Res* 1997;42:690-699. 939

79. du Plessis A. Cerebrovascular injury in premature infants: current 940  
understanding and challenges for future prevention. *Clin Perinatol* 941  
2008;35:609-641. 942

80. Soul J, Hammer P, Tsuji M, et al. Fluctuating pressure-passivity is 943  
common in the cerebral circulation of sick premature infants. 944  
*Pediatr Res* 2007;61:467-473. 945

81. Papile L, Rudolph AM, Heymann M. Autoregulation of cerebral 946  
blood flow in the preterm fetal lamb. *Pediatr Res* 1985;19:159-161. 947

82. Tweed W, Cote J, Pash M, Lou H. Arterial oxygenation deter- 948  
mines autoregulation of cerebral blood flow in fetal lamb. *Pediatr* 949  
*Res* 1985;17:246-249. 950

83. Helou S, Koehler RC, Gleason CA, Jones MD, Traystman RJ. 951  
Cerebrovascular Autoregulation During Fetal Development in 952  
Sheep. *Am J Physiol* 1994;266:H1069-H1074. 953

84. Hohimer AR, Bissonnette JM. Effects of cephalic hypotension, 954  
hypertension, and barbiturates on fetal cerebral blood flow and 955  
metabolism. *Am J Obstet Gynecol* 1989;161:1344-1351. 956

85. Greisen G. To autoregulate or not to autoregulate — that is no 957  
longer the question. *Semin Perinatol* 2009;16:207-215. 958

86. Tsuji M, Saul J, du Plessis A, et al. Cerebral intravascular oxy- 959  
genation correlates with mean arterial pressure in critically ill 960  
premature infants. *Pediatrics* 2000;106:625-632. 961

87. Raad RA, Tan WK, Bennet L, et al. Role of the cerebrovascular and 962  
metabolic responses in the delayed phases of injury after transient 963  
cerebral ischemia in fetal sheep. *Stroke* 1999;30:2735-2741. 964

88. Clapp III J, Peress N, Wesley M, Mann L. Brain damage after 965  
intermittent partial cord occlusion in the chronically instrumented 966  
fetal lamb. *Am J Obstet Gynecol* 1988;159:504-509. 967

89. Ikeda T, Murata Y, Quilligan E, et al. Physiologic and histologic 968  
changes in near-term fetal lambs exposed to asphyxia by partial 969  
umbilical cord occlusion. *Am J Obstet Gynecol* 1998;178:24-32. 970

90. Ohyu J, Marumo G, Ozawa H, et al. Early axonal and glial 971  
pathology in fetal sheep brains with leukomalacia induced by 972  
repeated umbilical cord occlusion. *Brain Dev* 1999;21:248-252. 973

91. Matsuda T, Okuyama K, Cho K, et al. Induction of antenatal 974  
periventricular leukomalacia by hemorrhagic hypotension in the 975  
chronically instrumented fetal sheep. *Am J Obstet Gynecol* 976  
1999;181:725-730. 977

92. Duncan J, Cock M, Scheerlinck J, et al. White matter injury after 978  
repeated endotoxin exposure in the preterm ovine fetus. *Pediatr* 979  
*Res* 2002;52:941-949. 980

93. Dalitz P, Harding R, Rees S, Cock M. Prolonged reductions in 981  
placental blood flow and cerebral oxygen delivery in preterm fetal 982  
sheep exposed to endotoxin: possible factors in white matter injury 983  
after acute infection. *J Soc Gynecol Investig* 2003;10:283-290. 984

94. Rees S, Stringer M, Just Y, Hooper S, Harding R. The vulnera- 985  
bility of the fetal sheep brain to hypoxemia at mid-gestation. 986  
*Devel Brain Res* 1997;103:103-118. 987

985 95. Mallard E, Rees S, Stringer M, Cock M, Harding R. Effects of chronic placental insufficiency on brain development in fetal sheep. *Pediatr Res* 1998;43:262-270.

986 96. Penning D, Grafe J, Hammond R, Matsuda Y, Patrick J, Richardson B. Neuropathology of the near-term and midgestation ovine fetal brain after sustained in utero hypoxemia. *Am J Obstet Gynecol* 1994;170:1425-1432.

987 97. Bagenholm R, Nilsson U, Gotborg C, Kjellmer I. Free radicals are formed in the brain of the fetal sheep during reperfusion after cerebral ischemia. *Pediatr Res* 1998;43:271-275.

988 98. Ikeda K, Murata Y, Quilligan EJ, Parer J, Doi S, Park S-D. Brain lipid peroxidation and antioxidant levels in fetal lambs 72 hours after asphyxia from partial umbilical cord occlusion. *Am J Obstet Gynecol* 1998;178:474-478.

989 99. Castillo-Melendez M, Chow J, Walker D. Lipid peroxidation, caspase-3 immunoreactivity, and pyknosis in late-gestation fetal sheep brain after umbilical cord occlusion. *Pediatr Res* 2004;55:864-871.

990 100. Welin A-K, Sandberg M, Lindblom A, et al. White matter injury following prolonged free radical formation in the 0.65 gestation fetal sheep brain. *Pediatr Res* 2005;58:100-105.

991 101. Fraser M, Bennet L, Van Zijl PL, et al. Extracellular amino acids and lipid peroxidation products in periventricular white matter during and after cerebral ischemia in preterm fetal sheep. *J Neurochem* 2008;105:2214-2223.

992 102. Baldwin B, Bell F. The anatomy of the cerebral circulation of the sheep and ox. The dynamic distribution of the blood supplied by the carotid and vertebral arteries to cranial regions. *J Anat* 1963;97:203-215.

993 103. Hohimer AR, Chao CR, Bissonnette JM. The effect of combined hypoxemia and cephalic hypotension on fetal cerebral blood flow metabolism. *xxxx* 1991;11:99-105.

994 104. Yoshimoto HS, Kaufman T, Keil LC, Rudolph AM. Responses to acute hypoxemia in fetal sheep at 0.6-0.7 gestation. *Am J Physiol* 1989;256(3 pt 2):H613-H620.

995 105. Rurak DW, Richardson BS, Patrick JE, Carmichael L, Homan J. Oxygen consumption in the fetal lamb during sustained hypoxemia with progressive acidemia. 1990;258(5 pt 2):R1108-R1115.

996 106. Richardson BS, Rurak D, Patrick JE, Homan J, Carmichael L. Cerebral oxidative metabolism during sustained hypoxaemia in fetal sheep. 1989;11:37-43.

997 107. Yan EB, Baburamani AA, Walker AM, Walker DW. Changes in cerebral blood flow, cerebral metabolites, and breathing movements in the sheep fetus following asphyxia produced by occlusion of the umbilical cord. *Am J Physiol Regul Integr Comp Physiol* 2009;297:R60-R69.

998 108. Richardson BS. The fetal brain: metabolic and circulatory responses to asphyxia. *Clin Invest Med* 1993;16:103-114.

999 109. McClure M, Riddle A, Manese M, et al. Cerebral blood flow heterogeneity in preterm sheep: lack of physiological support for vascular boundary zones in fetal cerebral white matter. *J Cereb Blood Flow Metab* 2008;28:995-1008.

1000 110. Chao CR, Hohimer AR, Bissonnette JM. Fetal cerebral blood flow and metabolism during oligemia and early postlogemic reperfusion. *J Cereb Blood Flow Metab* 1991;11:416-423.

1001 111. Nelson MD Jr., Gonzalez-Gomez I, Gilles FH, Dyke Award. The search for human telencephalic ventriculofugal arteries. 1991;12:215-222.

1002 112. Mayer PL, Kier EL. The controversy of the periventricular white matter circulation: a review of the anatomic literature. 1991;12:223-228.

1003 113. Volpe JJ. The structure of blood vessels in the germinal matrix and the autoregulation of cerebral blood flow in premature infants — reply. *Pediatrics* 2001;108:1050.

1004 114. Buser J, Segovia K, Dean J, et al. Timing of appearance of late oligodendrocyte progenitors coincides with enhanced susceptibility of preterm rabbit cerebral white matter to hypoxia-ischemia. *J Cereb Blood Flow Metab* 2010;30:1053-1065.

1005 115. Miller S, Ferrero D. From selective vulnerability to connectivity: insights from newborn brain imaging. *Trends Neurosci* 2009;32:496-505.

1006 116. Ment LR, Hirtz D, Huppi PS. Imaging biomarkers of outcome in the developing preterm brain. *Lancet Neurol* 2009;8:1042-1055.

1007 117. Mathur AM, Neil JJ, Inder TE. Understanding brain injury and neurodevelopmental disabilities in the preterm infant: the evolving role of advanced magnetic resonance imaging. *Semin Perinatol* 2010;34:57-66.

1008 118. Rutherford MA, Supramaniam V, Ederies A, et al. Magnetic resonance imaging of white matter diseases of prematurity. *Neuroradiology* 2010;52:505-521.

1009 119. Hope PL, Gould SJ, Howard S, Hamilton PA, Costello AM, Reynolds EO. Precision of ultrasound diagnosis of pathologically verified lesions in the brains of very preterm infants. *Dev Med Child Neurol* 1988;30:457-471.

1010 120. Schouman-Claeys E, Henry-Feugeas MC, Roset F, et al. Periventricular leukomalacia: correlation between MR imaging and autopsy findings during the first 2 months of life. *Radiology* 1993;189:59-64.

1011 121. Felderhoff-Mueser U, Rutherford MA, Squier WV, Cox P, Maalouf EF, Counsell SJ, et al. Relationship between MR imaging and histopathologic findings of the brain in extremely sick preterm infants. *AJNR Am J Neuroradiol* 1999;20:1349-1357.

1012 122. Inder TE, Neil JJ, Kroenke CD, Dieni S, Yoder B, Rees S. Investigation of cerebral development and injury in the prematurely born primate by magnetic resonance imaging and histopathology. *Dev Neurosci* 2005;27:100-111.

1013 123. Childs AM, Cornette L, Ramenghi LA, et al. Magnetic resonance and cranial ultrasound characteristics of periventricular white matter abnormalities in newborn infants. *Clin Radiol* 2001;56:647-655.

1014 124. Haynes RL, Billiards SS, Borenstein NS, Volpe JJ, Kinney HC. Diffuse axonal injury in periventricular leukomalacia as determined by apoptotic marker fractin. *Pediatr Res* 2008;63:656-661.

1015 125. Eklin S, Mallard C, Leverin A, et al. Bacterial endotoxin sensitizes the immature brain to hypoxic-ischaemic injury. *Eur J Neurosci* 2001;13:1101-1106.

1016 126. Eklin S, Mallard C, Arvidsson P, Hagberg H. Lipopolysaccharide induces both a primary and a secondary phase of sensitization in the developing rat brain. *Pediatr Res* 2005;58:112-116.

1017 127. Favrais G, van de Looij Y, Fleiss B, et al. Systemic inflammation disrupts the developmental program of white matter. *Ann Neurol* 2011;70:550-565.

1018 128. Filan PM, Hunt RW, Anderson PJ, Doyle LW, Inder TE. Neurologic Outcomes in Very Preterm Infants Undergoing Surgery. *J Pediatr* 2011;xx:xx-xx.

1019 129. Tam EW, Chau V, Ferrero D, et al. Preterm cerebellar growth impairment after postnatal exposure to glucocorticoids. *Sci Transl Med* 2011;3:105ra.

1020 130. Brambrink AM, Evers AS, Avidan MS, et al. Isoflurane-induced neuroapoptosis in the neonatal rhesus macaque brain. *Anesthesiology* 2010;112:834-841.

1021 131. Heine VM, Griveau A, Chapin C, Ballard PL, Chen JK, Rowitch DH. A small-molecule smoothened agonist prevents glucocorticoid-induced neonatal cerebellar injury. *Sci Transl Med* 2011;3:105ra4.

1022 132. Neubauer AP, Voss W, Kattner E. Outcome of extremely low birth weight survivors at school age: the influence of perinatal parameters on neurodevelopment. *Eur J Pediatr* 2008;167:87-95.

1023 133. Back SA, Luo NL, Borenstein NS, Levine JM, Volpe JJ, Kinney HC. Late oligodendrocyte progenitors coincide with the developmental window of vulnerability for human perinatal white matter injury. *J Neurosci* 2001;21:1302-1312.

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